combinatoria CHEMISTRY

Report

Subscriber access provided by American Chemical Society

Kinetic Comparison of Amide Formation on Various Cross-Linked Polystyrene Resins

Wenbao Li, Xiaoyi Xiao, and Anthony W. Czarnik

J. Comb. Chem., 1999, 1 (2), 127-129• DOI: 10.1021/cc980020I • Publication Date (Web): 11 February 1999

Downloaded from http://pubs.acs.org on March 20, 2009

More About This Article

Additional resources and features associated with this article are available within the HTML version:

- Supporting Information
- Access to high resolution figures
- Links to articles and content related to this article
- Copyright permission to reproduce figures and/or text from this article

View the Full Text HTML





© Copyright 1999 by the American Chemical Society

Volume 1, Number 2

March/April 1999

Kinetic Comparison of Amide Formation on Various Cross-Linked Polystyrene Resins

Wenbao Li,* Xiaoyi Xiao, and Anthony W. Czarnik[†]

ChemRx/IRORI, Discovery Partners International, 11149 North Torrey Pines Road, La Jolla, California 92037

Received September 23, 1998

Solid-phase organic synthesis (SPOS) is widely used to construct small molecule combinatorial libraries for drug discovery.¹ Advantages of SPOS over solution synthesis include the ease of product isolation and the ability to drive reactions to completion by the use of excess reagents. However, not all solution chemistry can be directly transferred to solid supports, and the conversion from solution chemistry to solid phase can be a slow, trial-and-error process, due in part to the lack of kinetic and mechanistic information for reactions on solid supports. Quantitative studies of solid-phase reactions can provide reaction rate and conversion data, as well as valuable information about reaction mechanisms. Recently, the kinetic effects of polystyrene (PS)- and TentaGel-based resins have been reported for several solid-phase organic reactions using single-bead IR and fluorescence methods.^{2–4} Here, we report a kinetic comparison of amide formation on various cross-linked polystyrene resins, using the attachment of Knorr linker as the model reaction (Scheme 1).

Figure 1 depicts reaction time courses of two independent amide formation experiments on 100–200 mesh aminomethyl PS resin using 0.2 M Knorr/0.4 M *N*,*N*-diisopropylethylamine (DIEA)/0.2 M benzotriazole-1-yl-oxy-tris-pyrrolidino-phosphonium hexafluorophosphate (PyBOP) in either dichloromethane (DCM) or dimethylformamide (DMF) at 25 °C. The amount of the reactive intermediates formed by the carboxylic acid and PyBOP^{5,6} is present in approximately 20 times molar excess over the amino groups on the resins. Therefore, the reaction can be treated as a pseudo-first-order reaction and be expressed as







Figure 1. Time courses of Knorr attachment onto 100–200 mesh aminomethyl polystyrene resin (0.5 mmol) in DCM (circles) and DMF (squares). Experimental conditions: 0.2 M Knorr, 0.4 M DIEA, 0.2 M PyBOP, 50 mL of DCM or DMF, 25 °C. Lines are the fitting of eq 1 with an observed rate constant of $2.1 \times 10^{-2} \text{ s}^{-1}$ and a reaction conversion of 93% in DCM, or with an observed rate constant of $7.9 \times 10^{-3} \text{ s}^{-1}$ and a reaction conversion of 71% in DMF.

Scheme 1



$$y = a_0 (1 - e^{-k_{obs}t})$$
 (1)

where k_{obs} is the observed reaction rate constant, a_0 is the amide formation conversion percentage when the reaction is completed, and *y* is the measured Knorr resin product percentage at each time point by Fmoc analysis.⁷ Then, the reaction half-life ($t_{1/2}$) can be calculated with eq 2.

$$t_{1/2} = 0.693/k_{\rm obs} \tag{2}$$

An observed rate constant of $2.1 \times 10^{-2} \text{ s}^{-1}$ and half-life ($t_{1/2}$) of 33 s were obtained on 100–200 mesh PS resin by

Table 1. Kinetics of Knorr Formation Reaction on Various $\operatorname{Resins}^{a}$

resins (size)	k _{obs} (1/s)	<i>t</i> _{1/2} (s)	con- version (%)	loading (mmol/g) ^b
PS (100-200 mesh)	2.1×10^{-2}	33	93	1.38
PS (100–200 mesh) ^c	7.9×10^{-3}	88	71	1.38
PS (200-400 mesh)	ca. 8 \times 10 ⁻²	ca. 9	96	0.43
PS (70-90 mesh)	3.9×10^{-3}	178	96	1.12
Champion-1	ca. 4×10^{-1}	ca. 2	98	0.40
$(100-200 \text{ mesh})^d$				
ArgoGel-AM ((164 μ m) ^e	ca. 7 \times 10 ⁻²	ca. 10	94	0.44
TentaGel-AM ((130 μ m) ^e	ca. 6 × 10 ⁻²	ca. 12	91	0.29

^{*a*} Experimental conditions: 0.2 M Knorr/0.4 M DIEA/0.2 M PyBOP in DCM at 25 °C. ^{*b*} Measured loading by Fmoc-Cl before Knorr formation (see Supporting Information for details). ^{*c*} The same experimental conditions used in previous entry but carried out in DMF instead of DCM. ^{*d*} Champion-1, a type of PEG-AMPS resin that has 60% PEG content from Biosearch Technologies.⁸ ^{*e*} Scheme 2.

fitting the duplicated data obtained using DCM (Figure 1 and Table 1). The reaction conversion is 93%, which has been corrected for the weight increase of the attached Knorr component on the aminomethyl resins (see Experimental Section for details).

When the same reaction was performed in DMF, an observed rate constant of $7.9 \times 10^{-3} \text{ s}^{-1}$ ($t_{1/2} = 88 \text{ s}$) and a corrected reaction conversion of 71% were fitted with eq 1 (Figure 1 and Table 1). The slower reaction rate and lower conversion could be at least partially explained by the lower swelling of PS resin in DMF as compared to that in DCM.

Table 1 also lists the kinetic constants for the same reaction using 200–400 and 70–90 mesh PS resins in DCM. The observed rate constants are ca. $8 \times 10^{-2} \text{ s}^{-1}$ and $3.9 \times 10^{-3} \text{ s}^{-1}$, respectively. The results indicate that the amide formation is faster on smaller (larger mesh) resins.

Also, we carried out the same kinetic study on 100-200 mesh poly(ethylene glycol) (PEG)-AMPS Champion-1 resin consisting of ~60% PEG and ~40% PS content. The reaction is very fast in DCM, with an observed rate constant of ca. 0.4 s⁻¹ (Table 1). The faster reaction rate on this PEG resin indicates that the PEG moiety plays an important and favorable role in the amide formation. Recently, excellent swelling properties and fast acylation characteristics of this resin have been reported.⁸

To further establish the effect of the PEG moiety in resins, two additional PEG-containing resins, namely ArgoGel-AM (~70% PEG-containing) and TentaGel-AM (70–80% PEGcontaining), were also studied under the identical conditions in DCM (Scheme 2). The observed rate constants are ca. 6 $\times 10^{-2}$ s⁻¹ and ca. 7 $\times 10^{-2}$ s⁻¹ for TentaGel and ArgoGel, respectively (Table 1). Unique properties of the solutionlike PEG long chains, such as favorable PEG-induced local environmental changes in solvation, dielectric properties, and hydrogen bonding, are possible explanations for the faster kinetics;⁹ however, this effect is highly reaction-dependent.^{3,4}

In summary, a kinetic comparison of amide formation on a variety of solid supports has been performed. Results indicate that amide formation is faster on smaller size (larger mesh) and PEG-containing resins, largely due to the resin size and the solution-like reaction environment.



Acknowledgment. We gratefully acknowledge valuable discussions with Drs. Shuhao Shi and Rongshi Li and support for the project from other colleagues at ChemRx/IRORI.

Supporting Information Available. Experimental Section. This information is available free of charge via the Internet at http://pubs.acs.org.

References and Notes

- (1) Jung, G.; Beck-Sickinger, A. G. Multiple Peptide Synthesis Methods and Their Applications. Angew. Chem., Int. Ed. Engl. 1992, 31, 367-386. (b) Pavia, M. R.; Sawyer, T. K.; Moos, W. H. The Generation of Molecular Diversity. Bioorg. Med. Chem. Lett. 1993, 3, 387-396. (c) Gordon, E. M.; Barrett, R. W.; Dower, W. J.; Fodor, S. P.; Gallop, M. A. Applications of Combinatorial Technologies to Drug Discovery. 2. Combinatorial Organic Synthesis, Library Screening Strategies, and Future Directions. J. Med. Chem. 1994, 37, 1385-1401. (d) Fruchtel, J. S.; Jung, G. Organic Chemistry on Solid Supports. Angew. Chem., Int. Ed. Engl. 1996, 35, 17-42. (e) Thompson, L. A.; Ellman, J. A. Synthesis and Application of Small Molecule Libraries. Chem. Rev. 1996, 96, 555-600. (f) DeWitt, D. H.; Czarnik, A. W. Combinatorial Organic Synthesis Using Parke-Davis's DIVERSOMER Method. Acc. Chem. Res. 1996, 29, 114-122. (g) Still, W. C. Discovery of Sequence-Selective Peptide Binding by Synthetic Receptors Using Encoded Combinatorial Libraries. Acc. Chem. Res. 1996, 29, 155-163. (h) Armstrong, R. W.; Combs, A. P.; Tempest, P. A.; Brown, S. D.; Keating, T. A. Multiple-Component Condensation Strategies for Combinatorial Library Synthesis. Acc. Chem. Res. 1996, 29, 123-131. (i) Ellman, J. A. Design, Synthesis, and Evaluation of Small-Molecule Libraries, Acc. Chem. Res. 1996, 29, 132-143. (j) Gordon, E. M.; Gallop, M. A.; Patel, D. V. Strategy and Tactics in Combinatorial Organic Synthesis: Application to Drug Discovery. Acc. Chem. Res. 1996, 29, 144-154. (k) Balkenhohl, F.; von Bussche-Hunnefeld, C.; Lansky, A.; Zechel, C. Combinatorial Synthesis of Small Organic Molecules. Angew. Chem., Int. Ed. Engl. 1996, 35, 2289-2337. (1) Lam, K. S.; Lebl, M.; Krchnak, V. The "One-Bead-One-Compound" Combinatorial Library Method. Chem. Rev. 1997, 97, 411-448. (m) Nefzi, A.; Ostresh, J. M.; Houghten, R. A. The Current Status of Heterocyclic Combinatorial Libraries. Chem. Rev. 1997, 97, 449-472. (n) Fenniri, H. Recent Advances at the Interface of Medicinal and Combinatorial Chemistry View on Methodologies for the Generation and Evaluation of Diversity and Application to Molecular Recognition and Catalysis. Current Med. Chem. 1996, 3, 343-378. (o) Brown, R. Future Pathways for Combinatorial Chemistry. Mol. Diversity 1997, 2, 217-222. (p) Lam, K. S. Application of Combinatorial Library Methods in Cancer Research and Drug Discovery. Anti-Cancer Drug Des. 1997, 12, 145 - 167
- (2) Yan, B.; Li, W. Rapid Fluorescence Determination of the Absolute Amount of Aldehyde and Ketone Groups on Resin Supports. J. Org. Chem. 1997, 62, 9354–9357.
- (3) Li, W.; Yan, B. Effects of Polymer Supports on the Kinetics of Solid-Phase Organic Reactions: A Comparison of Polystyrene- and TentaGel-Based Resins. J. Org. Chem. 1998, 63, 4092–4097.
- (4) Czarnik, A. W. Solid-Phase Synthesis Supports Are Like Solvents. Biotechnol. Bioeng. (Comb. Chem.) 1998, 61, 77–79.

Reports

- (6) Hudson, D. Methodological Implications of Simultaneous Solid-Phase Peptide Synthesis. 1. Comparison of Different Coupling Procedures. *J. Org. Chem.* **1988**, *53*, 617–624.
- (7) Fields, G. B.; Noble, R. L. Solid-Phase Peptide Synthesis Utilizing Fluorenylmethyloxycarbony Amino Acids. Int. J. Peptide Protein Res. 1990, 35, 161–214.

- (8) Adams, J. H.; Cook, R. M.; Hudson, D.; Jammalamadaka, V.; Lyttle, M.; Songster, M. F. A Reinvestigation of the Preparation, Properties, and Applications of Aminomethyl and 4-Methylbenzhydrylamine Polystyrene Resins. J. Org. Chem. **1998**, 63, 3706–3716.
- (9) Mutter, M.; Bayer, E. The Liquid-Phase Method for Peptide Synthesis. In *The Peptides: Analysis, Synthesis, Biology*; Gross, E., Meienhofer, J., Eds.; Academic Press: New York, 1979; Vol. 2, pp 285–332.

CC980020L